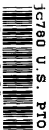


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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of: Kristian Berg et al.

Title: METHOD OF VACCINATION

Attorney Docket No.: 697.013US1

PATENT APPLICATION TRANSMITTAL

BOX PATENT APPLICATION

Assistant Commissioner for Patents

Washington, D.C. 20231

We are transmitting herewith the following attached items and information (as indicated with an "X"):

- X** Utility Patent Application under 37 CFR § 1.53(b) comprising:
- X** Specification (28 pgs, including claims numbered 1 through 21 and a 1 page Abstract).
 - X** Formal Drawing(s) (5 sheets).
 - X** Unsigned Combined Declaration and Power of Attorney (4 pgs).
 - X** Preliminary Amendment (2 pgs).
 - X** Return postcard.

The filing fee (NOT ENCLOSED) will be calculated as follows:

	No. Filed	No. Extra	Rate	Fee
TOTAL CLAIMS	21 - 20 =	1	x 9 =	\$9.00
INDEPENDENT CLAIMS	9 - 3 =	6	x 39 =	\$234.00
MULTIPLE DEPENDENT CLAIMS PRESENTED				\$0.00
BASIC FEE				\$345.00
TOTAL				\$588.00

THE FILING FEE WILL BE PAID UPON RECEIPT OF THE NOTICE TO FILE MISSING PARTS.

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S/N Unknown

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: Kristian Berg et al.

Examiner: Unknown

Serial No.: Unknown

Group Art Unit: Unknown

Filed: Herewith

Docket: 697.013US1

Title: METHOD OF VACCINATION (as amended)

PRELIMINARY AMENDMENT

BOX PATENT APPLICATION

Assistant Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Prior to examination of the above-identified patent application please amend as follows:

IN THE TITLE

Please delete "METHOD" and insert --METHOD OF VACCINATION-- therefor.

IN THE CLAIMS

In claim 3, line 1, please delete "or claim 2".

In claim 5, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 6, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 7, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 9, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 10, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 11, line 1, please delete "any of the preceding claims" and insert --claim 1-- therefor.

In claim 12, line 2, please delete "any of the preceding claims" and insert --claim 1-- therefor.

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PRELIMINARY AMENDMENT

Serial Number: Unknown

Filing Date: Herewith

Title: PCI VACCINATION

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Dkt: 697.013US1

In claim 13, line 4, please delete "any one of claims 1 to 5 or claim 9" and insert --claim 1-- therefor.

In claim 13, line 5-6, please delete "any one of claims 2 or 7 to 9" and insert --claim 2-- therefor.

In claim 14, line 4, please delete "in any one of claims 1 to 12" and insert --claim 1-- therefor.

In claim 16, line 2, please delete "any of the preceding claims" and insert --claim 2-- therefor.

In claim 18, line 1, please delete "or claim 17".

In claim 19, line 1, please delete "any one of claims 16 to 18" and insert --claim 16-- therefor.

In claim 20, line 2, please delete "any one of claims 1 to 5 or claim 9" and insert --claim 1-- therefor.

In claim 20, lines 3-4, please delete "any one of claims 2 or 7 to 9" and insert --claim 2-- therefor.

In claim 21, lines 5-6, please delete "any one of claims 1 to 5 or claim 9" and insert --claim 1-- therefor.

In claim 21, line 8, please delete "any one of claims 2 or 7 to 9" and insert --claim 2-- therefor.

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Washington, D.C. 20231

Printed Name Shawn Hise

Signature [Signature]

Respectfully submitted,

KRISTIAN BERG ET AL.

By their Representatives,

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Date March 10, 2000

By Ann M. McCrackin
Ann M. McCrackin
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METHOD

5 The present invention relates to a method of
vaccination which involves using photodynamic treatment
(PDT) to introduce vaccine components into cells to
achieve antigen presentation, and to vaccine
compositions useful in such a method.

10 The majority of molecules do not readily penetrate
cell membranes. Methods for introducing molecules into
the cytosol of living cells are useful tools for
manipulating and studying biological processes. Among
the most commonly used methods today are microinjection,
red blood cell ghost-mediated fusion and liposome
15 fusion, osmotic lysis of pinosomes, scrape loading,
electroporation, calcium phosphate and virus-mediated
transfection. These techniques are useful for
investigating cells in culture, although in many cases
they may be impractical, time consuming, inefficient or
20 they may induce significant cell death. Thus such
techniques are not optimal for use in biological or
medical research, or in therapies, where it is required
that cells should remain viable and/or functional.

25 It is well known that porphyrins and many other
photosensitizing compounds may induce cytotoxic effects
on cells and tissues. These effects are based upon the
fact that upon exposure to light the photosensitizing
compound may become toxic or may release toxic
substances such as singlet O_2 or other oxidising radicals
30 which are damaging to cellular material or biomolecules,
including the membranes of cells and cell structures,
and such cellular or membrane damage may eventually kill
the cells. These effects have been utilised in the
treatment of various abnormalities or disorders,
35 including especially neoplastic diseases. The treatment
is named photodynamic therapy (PDT) and involves the
administration of photosensitizing

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(photochemotherapeutic) agents to the affected area of the body, followed by exposure to photoactivating light in order to activate the photosensitizing agents and convert them into cytotoxic form, whereby the affected cells are killed or their proliferative potential diminished. Photosensitizing agents are known which will localise preferentially or selectively to the desired target site e.g. to a tumour or other lesion.

A range of photosensitizing agents are known, including notably the psoralens, the porphyrins, the chlorins and the phthalocyanins. Such drugs become toxic when exposed to light.

Photosensitizing drugs may exert their effects by a variety of mechanisms, directly or indirectly. Thus for example, certain photosensitisers become directly toxic when activated by light, whereas others act to generate toxic species, e.g. oxidising agents such as singlet oxygen or other oxygen-derived free radicals, which are extremely destructive to cellular material and biomolecules such as lipids, proteins and nucleic acids.

Porphyrin photosensitisers act indirectly by generation of toxic oxygen species, and are regarded as particularly favourable candidates for PDT. Porphyrins are naturally occurring precursors in the synthesis of heme. In particular, heme is produced when iron (Fe^{2+}) is incorporated in protoporphyrin IX (Pp) by the action of the enzyme ferrochelatase. Pp is an extremely potent photosensitizer, whereas heme has no photosensitizing effect. A variety of porphyrin-based or porphyrin-related photosensitisers are known in the art and described in the literature.

The cytotoxic effect is mediated mainly through the formation of singlet oxygen. This reactive intermediate has a very short lifetime in cells ($<0.04 \mu\text{s}$). Thus, the primary cytotoxic effect of PDT is executed during light exposure and very close to the sites of formation of $^1\text{O}_2$. $^1\text{O}_2$ reacts with and oxidizes proteins (histidine,

tryptophan, methionine, cysteine, tyrosine), DNA (guanine), unsaturated fatty acids and cholesterol. One of the advantages of PDT is that tissues unexposed to light may be left unaffected ie. that a selective PDT effect may be obtained. There is extensive documentation regarding use of PDT to destroy unwanted cell populations, for example neoplastic cells. The patent literature describes a number of photodynamic compounds, alone or conjugated with targeting agents, e.g. immunoglobulins directed to neoplastic cell receptor determinants, making the complex more cell specific. Certain photochemical compounds, such as hematoporphyrin derivatives, have furthermore an inherent ability to localise in malignant cells. Such methods and compounds, are described in the Norwegian patent NO 173319, in Norwegian patent applications Nos. 90 0731, 176 645, 176 947, 180 742, 176 786, 301 981, 30 0499 and 89 1491.

In WO93/14142 a drug delivery system is described which comprises an anti-cancer agent and a photoactivatable agent (ie. a photosensitizer) attached to copolymeric carriers. Upon administration this complex enters the cell interior by pinocytosis or phagocytosis and locates inside the endosomes and lysosomes. In the lysosomes, the bond between the anti-neoplastic agent and the polymer is hydrolysed and the former can diffuse passively through the lysosome membrane into the cytosol. The utility of this method is thus limited to small molecular compounds which are able to diffuse across the lysosome membranes. After allowing a time lag for diffusion, a light source of appropriate wavelength and energy is applied to activate the photo-activatable compound. The combined effect of the anti-cancer agent and photoactivatable agent destroy the cell.

Such PDT methods as described above are thus directed to the destruction of cell structures leading

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to cell death.

WO 96/07432, on the other hand, is concerned with methods which use the photodynamic effect as a mechanism for introducing otherwise membrane-impermeable molecules into the cytosol of a cell in a manner which does not result in widespread cell destruction or cell death. In this method, the molecule is co-internalised (more particularly "endocytosed") into an intracellular vesicle in the cell (e.g. a lysosome or endosome) together with a photosensitizing agent. The cell is then exposed to photoactivating light which "activates" the photosensitizer, which in turn causes the vesicle membrane to disrupt or rupture, releasing the vesicle contents, including the molecule, into the cell interior ie. the cytosol. It was found that in such a method the functionality or the viability of the majority of the cells was not deleteriously affected. Thus, the utility of such a method, termed "photochemical internalisation" was proposed for transporting a variety of different molecules, including therapeutic agents, into the cytosol ie. into the interior of a cell.

We have now found that such a method can advantageously be used, not only to transfer molecules in the interior of a cell, but also to present or express them on a cell surface. Thus, following transport and release of a molecule into the cell cytosol, it may be transported to the surface of the cell where it may be presented on the outside of the cell ie. on the cell surface. Such a method has particular utility in the field of vaccination, where vaccine components ie. antigens or immunogens, may be introduced to a cell for presentation on the surface of that cell, in order to induce, facilitate or augment an immune response.

At its most general, the present invention thus provides a method of expressing an antigenic molecule or a part thereof on the surface of a cell, preferably an

antigen-presenting cell, said method comprising
introducing a molecule into the cell cytosol by
photochemical internalisation, wherein said molecule, or
a part thereof, is subsequently presented on the surface
of said cell.

As used herein "expressing" or "presenting" refers
to the presence of the molecule or a part thereof on the
surface of said cell such that at least a portion of
that molecule is exposed and accessible to the
environment surrounding that cell. Expression on the
"surface" may be achieved in which the molecule to be
expressed is in contact with the cell membrane and/or
components which may be present or caused to be present
in that membrane.

Such antigenic presentation may advantageously
result in the stimulation of an immune response,
preferably an immune response which confers protection
against subsequent challenge by an entity comprising or
containing said antigen molecule or part thereof, and
consequently the invention finds particular utility as a
method of vaccination.

More particularly, this aspect of the invention
provides a method of expressing an antigenic molecule or
a part thereof on the surface of a cell, said method
comprising:

contacting said cell with said antigenic molecule
and with a photosensitizing agent, wherein said molecule
and said agent are each taken up into an intracellular
membrane-restricted compartment of said cell; and

irradiating said cell with light of a wavelength
effective to activate the photosensitizing agent, such
that the membrane of said intracellular compartment is
disrupted, releasing said molecule into the cytosol of
the cell, without killing the cell,

wherein, said released antigenic molecule, or a
part thereof, is subsequently presented on the surface
of said cell.

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As used herein, a "disrupted" compartment refers to destruction of the integrity of the membrane of that compartment either permanently or temporarily, sufficient to allow release of the antigenic molecule contained within it.

Alternatively viewed, this aspect of the invention also provides a composition for use in expressing an antigenic molecule or a part thereof on the surface of a cell, preferably to simulate an immune response, said composition comprising an antigenic molecule and a photosensitizing agent. Preferably said composition is pharmaceutically acceptable and contains also a pharmaceutically acceptable excipient or diluent.

In a further aspect, the invention also provides the use of an antigenic molecule and a photosensitizing agent in the preparation of a medicament for use in expressing said antigenic molecule or a part thereof on the surface of a cell to stimulate an immune response.

A still further aspect of the invention provides a product comprising an antigenic molecule and a photosensitizing agent as a combined preparation for simultaneous, separate or sequential use in expressing said antigenic molecule or a part thereof on the surface of a cell, preferably to stimulate an immune response.

A yet further aspect of the invention provides a kit for use in expressing an antigenic molecule or a part thereof on the surface of a cell, said kit comprising

a first container containing said antigenic molecule; and

a second container containing a photosensitizing agent.

In the invention, the antigenic molecule may be any molecule wherein that molecule or a part thereof is capable of stimulating an immune response, when presented to the immune system in an appropriate manner. Advantageously, therefore the antigenic molecule will be

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a vaccine antigen or vaccine component, such as a polypeptide containing entity.

Many such antigens or antigenic vaccine components are known in the art and include all manner of bacterial or viral antigens or indeed antigens or antigenic components of any pathogenic species including protozoa or higher organisms. Whilst traditionally the antigenic components of vaccines have comprised whole organisms (whether live, dead or attenuated) ie. whole cell vaccines, in addition sub-unit vaccines, ie. vaccines based on particular antigenic components of organisms e.g. proteins or peptides, or even carbohydrates, have been widely investigated and reported in the literature. Any such "sub-unit"-based vaccine component may be used as the antigenic molecule of the present invention. However, the invention finds particular utility in the field of peptide vaccines. Thus, a preferred antigenic molecule according to the invention is a peptide (which is defined herein to include peptides of both shorter and longer lengths ie. peptides, oligopeptides or polypeptides, and also protein molecules or fragments thereof e.g. peptides of 5-500 e.g. 10 to 250 such as 15 to 75, or 8 to 25 amino acids). Parts of antigenic molecules which are presented or expressed preferably comprise parts which are generated by antigen-processing machinery within the cell. Parts may however be generated by other means which may be achieved through appropriate antigen design (e.g. pH sensitive bands) or through other cell processing means. Conveniently such parts are of sufficient size to generate an immune response, e.g. in the case of peptides greater than 5, e.g. greater than 10 or 20 amino acids in size.

A vast number of peptide vaccine candidates have been proposed in the literature, for example in the treatment of viral diseases and infections such as AIDS/HIV infection or influenza, canine parvovirus, bovine leukaemia virus, hepatitis, etc. (see e.g. Phanuphak et

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in the immune response. However, other cells may also present antigen to the immune system and these also fall within the scope of the invention. The cells according to the present invention are thus advantageously antigen-presenting cells. The antigen-presenting cell may be involved in any aspect or "arm" of the immune response, including both humoral and cell-mediated immunity, for example the stimulation of antibody production, or the stimulation of cytotoxic or killer cells, which may recognise and destroy (or otherwise eliminate) cells expressing "foreign" antigens on their surface. The term "stimulating an immune response" thus includes all types of immune responses and mechanisms for stimulating them.

The stimulation of cytotoxic cells or antibody-producing cells, requires antigens to be presented to the cell to be stimulated in a particular manner by the antigen-presenting cells, for example MHC Class I presentation (e.g. activation of CD8⁺ cytotoxic T-cells requires MHC-1 antigen presentation).

Antigen-presenting cells are known in the art and described in the literature and include for example, lymphocytes (both T and B cells), dendritic cells, macrophages etc. Others include for example cancer cells e.g. melanoma cells.

For antigen presentation by an antigen-presenting cell to a cytotoxic T-cell (CTL) the antigenic molecule needs to enter the cytosol of the antigen-presenting cell (Germain, Cell, 1994, 76, 287-299). The present invention provides an efficient means of delivery of the antigenic molecule into the cytosol.

Once released in the cell cytosol by the photochemical internalisation process, the antigenic molecule may be processed by the antigen-processing machinery of the cell and presented on the cell surface in an appropriate manner e.g. by Class I MHC. This processing may involve degradation of the antigen, e.g.

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degradation of a protein or polypeptide antigen into peptides, which peptides are then complexed with molecules of the MHC for presentation. Thus, the antigenic molecule expressed or presented on the surface of the cell according to the present invention may be a part or fragment of the antigenic molecule which is internalised (endocytosed).

Antigens may be taken up by antigen-presenting cells by endocytosis and degraded in the endocytic vesicles to peptides. These peptides may bind to MHC class II molecules in the endosomes and be transported to the cell surface where the peptide-MHC class II complex may be recognised by CD4+ T helper cells and induce an immune response. Alternatively, proteins in the cytosol may be degraded, e.g. by proteasomes and transported into endoplasmic reticulum by means of TAP (transporter associated with antigen presentation) where the peptides may bind to MHC class I molecules and be transported to the cell surface as illustrated in the figure 1 (Yewdell and Bennink, 1992, Adv. Immunol. 52: 1-123). If the peptide is of foreign antigen origin, the peptide-MHC class I complex will be recognised by CD8+ cytotoxic T-cells (CTLs). The CTLs will bind to the peptide-MHC (HLA) class I complex and thereby be activated, start to proliferate and form a clone of CTLs. The target cell and other target cells with the same peptide-MHC class I complex on the cells surface may be killed by the CTL clone. Immunity against the foreign antigen may be established if a sufficient amount of the antigen can be introduced into the cytosol (Yewdell and Bennink, 1992, *supra*; Rock, 1996, Immunology Today 17: 131-137). This is the basis for development of *inter alia* cancer vaccines. One of the largest practical problems is to introduce sufficient amounts of antigens (or parts of the antigen) into the cytosol. This may be solved according to the present invention by PCI. This principle is illustrated in Fig.

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1, which shows how PCI can be utilised to stimulate CTLs. A peptide or protein (P) is applied extracellularly to antigen-presenting cells. P is endocytosed and released into cytosol by PCI. The peptide or protein will thereafter be partly degraded by proteasomes and transported to the cells surface complexed to MHC (HLA) class I where the complex can be recognised by CTLs.

As will be described in more detail in the Examples below, it has been demonstrated that photochemical internalisation may be used efficiently according to the present invention for cytosolic delivery of cancer-specific peptides.

The antigenic molecule and/or photosensitivity agent may be targeted to specific cells or tissues by employing targeting agents e.g. target-specific delivery or carrier systems or carrier molecules. Thus for example the antigenic molecule and/or photosensitising agent may be delivered to the cell using a vector or carrier system e.g. reconstituted LDL-particles. The carrier molecule may be bound or conjugated to the antigenic molecule, to the photosensitising agent or both, and the same or different carrier molecules may be used. The antigenic molecule and/or photosensitising agent may also be conjugated to a site-targeting ligand, such as a ligand which is specific for particular cell-types or particular cell structures e.g. an antibody recognising a surface antigen expressed on certain cell types e.g. a tumour-specific antigen. Such mechanisms may act to increase uptake of the photosensitiser and/or antigen molecule through receptor-mediated endocytosis. Such targeting molecules carriers or vectors may also be used to direct the antigenic molecule and/or photosensitising agent to the intracellular compartment.

The intracellular membrane-restricted compartment may be any such compartment which is present in a cell. Preferably the compartment will be a membrane vesicle,

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especially an endosome or a lysosome. However, the intracellular compartment may also include the Golgi apparatus or the endoplasmic reticulum. All that is required is that the antigenic molecule and the photosensitising agent locate to the same intracellular compartment(s).

The photochemical internalisation process is described in more detail in WO 96/07432 (the contents of which are incorporated herein by reference). Methods of PDT are also now widely described in the literature.

The photosensitizing agent to be used according to the present invention may be any such agent which localises to intracellular compartments, particularly endosomes or lysosomes. A range of such photosensitising agents are known in the art and are described in the literature, including in WO96/07432. Mention may be made this respect of di- and tetrasulfonated aluminium phthalocyanine, sulfonated tetraphenylporphines (TPPS_n), nile blue, chlorin e₆ derivatives, uroporphyrin I, phylloerythrin, hematoporphyrin and methylene blue which have been shown to locate in endosomes and lysosomes of cells in culture. This is in most cases due to endocytic activity.

Classes of suitable photosensitising agent which may be mentioned thus include porphyrins, phthalocyanines, purpurins, chlorins, benzoporphyrins naphthalocyanines, cationic dyes, tetracyclines and lysomotropic weak bases or derivatives thereof (Berg et al., Photochemistry and Photobiology, 1997, 65, 403-409).

Preferred photosensitising agents include TPPS₄, (Zabner et al., J. Biol. Chem. 1995, 270, 18997-19007) TPPS_{2a} and ALPcS_{2a}.

The photochemical internalisation according to the present invention may be carried out using PDT methods which are known and standard in the art and appropriate

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Thus, the compositions of the invention may be formulated in any convenient manner according to techniques and procedures known in the pharmaceutical

art, e.g. using one or more pharmaceutically acceptable carrier or excipients. The nature of the composition and carriers or excipient materials, dosages etc. may be selected in routine manner according to choice and the desired route of administration, purpose of vaccination etc. Dosages may likewise be determined in routine manner and may depend upon the nature of the antigenic molecule, purpose of vaccination, age of patient, mode of administration etc., in connection with the photosensitising agent the potency/ability to disrupt membranes on irradiation, should also be taken into account.

The light irradiation step to activate the photosensitising agent may likewise take place according to techniques and procedures well known in the art. For example, the wavelength and intensity of the light may be selected according to the photosensitising agent used. Suitable light sources are well known in the art.

As mentioned earlier, and as described in WO96/07432, it has been found that photochemical internalisation in this manner does not deleteriously affect the viability and functionality of the cells. In particular, it has been found that when a population or plurality of cells is treated according to the present invention, a majority of the cells are not killed, and survive the treatment, substantially functionally intact.

As used herein, the term "without killing the cell" is intended to define such a situation. In other words in a population or plurality of cells, substantially all of the cells, or a significant majority (e.g. at least 75%, more preferably at least 80, 85, 90 or 95% of the cells) are not killed.

Clearly when dealing with the light irradiation of a population or a plurality of cells it is possible that certain groups of cells or certain areas of tissue may receive more light or in some other way be subjected to

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that confer tissue specificity or the use of disease-specific drugs, opening a possibility of obtaining substantially synergistic effects in the specificity of drugs for target cells.

5 The invention will now be described in more detail in the following non-limiting Examples with reference to the following drawings in which:

Figure 1 shows a schematic representation of how PCI can be utilised to stimulate CTLs. A peptide or protein (P) is applied extracellularly to antigen
10 presenting cells. P is endocytosed and released into cytosol by PCI. The peptide or protein will thereafter be partly degraded by proteasomes and transported to the cells surface complexed to MHC (HLA) class I where the
15 complex can be recognised by CTLs.

Figure 2 shows photochemically induced relocalisation of a peptide. BL2-G-E6 cells were incubated with a fluorescein-labelled p21^{ras}-derived 5-21, Val¹² peptide and ALPcS_{2a}. The cells were examined
20 for fluorescein-peptide and ALPcS_{2a} localisation by fluorescence microscopy before (top panels) and 30 minutes after (bottom panels) a 4-min exposure to red light. Bar 20 μ m.

Figure 3 shows the cytotoxicity of a CD8⁺ T lymphocyte clone against FM3 melanoma cells after PCI of
25 a MART-1 peptide.

Figure 4 shows the ability of PCI to deliver HRP into the cytosol. NHIK 3025 cells were treated with 3.2 μ g/ml TPPS_{2a} and 1 mg/ml HRP for 18 hours. The medium
30 was then replaced with drug-free medium before exposure to the indicated light doses. HRP activity was measured in intact cells (○) and in cytosol (●) separated from cytosol-free cell corpses (▼) by electroporabilisation and a density centrifugation technique.

35 Figure 5 shows photochemically induced expression of GFP. a. expression of GFP in THX cells treated with PEGFP-N1-plys complex in the absence of ALPcS_{2a} and light

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or in the presence of AlPCS_{2a} followed by exposure to light as indicated on the figure. The cells were analysed by flow cytometry, reckoning the cells on the right side of the drawn line as positive for GFP

5 expression. b. expression of GFP in THX cells treated for 18 hours with a photosensitiser (20 µg/ml AlPCS_{2a} or 0.25 µg/ml 3-THPP) followed by a 6 hour transfection with pEGF-N1-pLys complex and exposure to light inactivating 50% of the cells. GFP expression was

10 analysed by flow cytometry as described in a.

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EXAMPLES

Materials and Methods

5 Irradiation

Two different light sources were used for treatment of the cells, both consisting of a bank of 4 fluorescent tubes. Cells treated with TPPS₁, TPPS_{2a}, and 3-THPP (Porphyrin Products, Logan, UT) were exposed to blue light (model 3026; Appl. Photophysics, London, UK) with a light intensity reaching the cells of 1.5 mW/cm² while cells treated with AlPcS_{2a} (Porphyrin Products, Logan, UT) were exposed to red light (Philips TL 20W/09) filtered through a Cinemoid 35 filter with a light intensity reaching the cells of 1.35 mW/cm².

Fluorescence microscopy

20 The cells were analysed by fluorescence microscopy as described in Berg. K., et al., Biochem. Biophys. Acta., 1370: 317-324, 1998. For analysis of fluorescein-labelled molecules the microscope was equipped with a 450-490 nm excitation filter, a 510 nm dichroic beam splitter and a 510-540 nm band pass emission filter.

Preparation of Plasmid-pLys Complexes and Treatment of cells

30 Plasmid-pLys complexes (charge ratio, 1.7) were prepared by gently mixing 5 µg plasmid (pEGFP-N1; Clontech Laboratories, Inc., Palo Alto, CA) in 75 µl of HBS with 5.3 µg pLys (MW 20700; Sigma, St. Louis, MO) in 75 µl of HBS. The solutions were incubated for 30 min at room temperature, diluted with culture medium and added to the cells.

THX cells were incubated with 20 $\mu\text{g/ml}$ AlPcS_{2a} for 18 hours at 37°C, washed and incubated in sensitizer-free medium for 3 hours before incubation with plasmid-pLys complexes for 2 hours. The pEGFP-N1/pLys treated THX cells were washed once and incubated for 2 hours in culture medium without additions before exposure to light. The cells were incubated at 37°C for 2 days, subcultured and further incubated for an additional 5 days before analysis of GFP expression by flow cytometry.

HCT-116 cells were incubated with 20 $\mu\text{g/ml}$ AlPcS_{2a} for 18 hours, washed and transfected with plasmid-pLys complexes for 6 hours before light exposure in plasmid-free medium. After 40 hours incubation at 37°C the GFP expression was studied by microscopy.

Flow cytometry analysis

The cells were trypsinised, centrifuged, resuspended in 400 μl of culture medium and filtered through a 50 μm mesh nylon filter. The cells were then analysed in a FACStar plus flow cytometer (Becton Dickinson). Green Fluorescent Protein (GFP) was measured through a 510-530 nm filter after excitation with an argon laser (200 mW) tuned on 488 nm. AlPcS_{2a} was measured through a 650 nm longpass filter after excitation with a krypton laser (50 mW) tuned on 351-356 nm. Cell doublets were discriminated from single cells by gating on the pulse width of the GFP fluorescence signal. The data were analysed with PC Lysys II software (Becton Dickinson).

Preparation of Fluorescein-Peptide and Treatment of Cells

The fluorescein-labelled Val¹²-p21^{ras}-peptide (residues 5-21) were synthesised and provided by Alan Cuthbertson,

Nycomed Amersham).

BL2-G-E6 cells were incubated with 30 $\mu\text{g}/\text{ml}$ of the
fluorescein-labelled p21^{ras}-derived peptide for 18 hours
5 followed by 20 $\mu\text{g}/\text{ml}$ AlPcS_{2a} for 18 hours and 1 hour in
drug-free medium before exposure to red light.

Example 1

10 Photochemical internalisation (PCI) can be used to
enable peptides to enter the cytosol of cells

To evaluate PCI for cytosolic delivery of cancer-
specific peptides, a fluorescein-labelled p21^{ras} peptide
15 encompassing residues 5-21 and containing a Val¹²
mutation (G12V) was used (Gjertsen, M.K., et al., Int.
J. Cancer, 72: 784-790, 1997). In BL2-6-E6 mouse
fibroblasts, the ras peptide colocalised well with
AlPcS_{2a}, indicating endocytic uptake of the peptide (Fig.
20 2). After a 4-min exposure to light, the fluorescein-
labelled ras peptide and AlPcS_{2a} were found to be located
diffusely in the cytoplasm. Similar effects were not
observed in cells exposed to the fluorescein-labelled
ras peptide and light only (data not shown).

25

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Example 2

Use of PCI to induce antigen presentation and CD8⁺ T lymphocyte mediated cell killing

5 FM3 melanoma cells (2×10^5 /well in 6 well plates), grown in RPMI 1640 medium with 10% foetal calf serum (FCS), not expressing MART-1 peptide were treated with 10 $\mu\text{g/ml}$ of the photosensitising agent ALPcS₂ for 18 hours. The
10 cells were then released from the substratum with EDTA (0.1 M) in Dulbecco's phosphate-buffered saline (PBS) and kept in solution during loading of the cells with ^{51}Cr (60 $\mu\text{Ci/ml}$ Na_2CrO_4) for 1 hour in 100% FCS followed by 5 hours incubation with 5 $\mu\text{g/ml}$ MART-1 peptide in
15 RPMI 1640 in 10% FCS, while the cells were still kept in solution. The sequence of the MART-1 peptide was: TABEAAGIGILTIVILG. The cells were then washed twice in RPMI 1640 medium containing 10% FCS and seeded out in
20 96-well plates (2000/well in 100 μl medium (RPMI 1640/10% FCS)). The cells were then exposed to light for the times as indicated in Figure 3 ((Philips TL 20W/09) filtered through a Cinemoid 35 filter with a light intensity reaching the cells of 1.35 mW/cm^2 (Rodal et al., 1998, J. Photochem. Photobiol. B: Biol. 45: 150-
25 9)). 18 hours after light exposure the medium was removed and medium containing MART-1/HLA-A2 specific cytotoxic T lymphocytes (CTLs - 40,000/well added in 100 μl) were added. After 4 hours of incubation the medium was separated from FM3 cells and the ^{51}Cr released to the
30 medium (as an indicator of lysed cells) was counted as well as the spontaneous and maximum release as previously described (Fossum et al., 1995, Cancer Immunol. Immunother. 40: 165-172). The percentage specific chromium release was calculated by the formula:
35 (experimental release - spontaneous release)/(maximum release - spontaneous release) x 100. It can be seen from the results shown in Figure 3 that FM3 cells after

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PCI of a MART-1 peptide as outlined above show light dependent susceptibility to CD8⁺ T lymphocyte cytotoxicity.

5 Example 3

PCI induces the release of a large fraction of the endocytosed molecule

10 This was shown by PCI induced internalisation/ endocytosis of Horseradish Peroxidase (HRP).

15 By using HRP, it is demonstrated (see Figure 4) that PCI induces the release of a large fraction (>60%) of endocytosed HRP into the cytosol.

20 In this experiment NHIK 3025 cells (carcinoma cells in situ from human cervix) were treated with the photosensitising agent TPPS_{2a} (3.2 µg/ml) and 1 mg/ml HRP for 18 hours. The medium was then replaced with drug free medium before exposure to the light doses as indicated in Figure 4. HRP activity was measured according to the procedure described in Steinman et al., J. Cell Biol., 68: 665-687, 1976. Cytosol was separated from cytosol-free cell corpses by
25 electropermeabilisation and a density centrifugation technique (Berg et al., Int. J. Cancer 59: 814-822, 1994).

30 Example 4

PCI can be used to enhance the delivery of functional genes

35 To demonstrate this, THX cells were transfected with a pLys-complex of a plasmid (pEGFP-N1) coding for green fluorescent protein (GFP). The expression of GFP was

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analysed by flow cytometry (Fig. 5, a and b) and
fluorescence microscopy (data not shown). As can be
seen from Fig. 5a, treatment with AlPCs_{2a} and light led
to a strong increase in the percentage of the cells
expressing GFP. The fraction of the cells that was
positive for this reporter molecule increased from 1% at
no light treatment to 50% after a 5-min light exposure.
GFP expression was not enhanced by light in cells
treated with pEGFP-pLys in the absence of a
photosensitiser. A complex of an irrelevant plasmid
(encoding heme oxygenase) and pLys did not induce green
fluorescence when combined with AlPCs_{2a} and light (data
not shown). Consequently, in a light-directed manner,
PCI can substantially increase the efficiency of
transfection of a functional gene to THX cells. Similar
results were obtained using TPPS_{2a} as a photosensitiser
and BHK-21 and HCT-116 as target cells (data not shown).
The essentially non-lysosomally located sensitiser 3-
THPP induced only a minor increase in GFP expression
(Fig. 5b). PCI of pEGFP-N1 not complexed with pLys did
not induce the expression of GFP (data not shown).

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Claims

1. A method of expressing an antigenic molecule or a part thereof on the surface of a cell, said method comprising introducing a molecule into the cell cytosol by photochemical internalisation, wherein said molecule, or a part thereof, is subsequently presented on the surface of said cell.

2. A method as claimed in claim 1, said method comprising:

contacting said cell with said antigenic molecule or a part thereof and with a photosensitizing agent, wherein said molecule and said agent are each taken up into an intracellular membrane-restricted compartment of said cell; and

irradiating said cell with light of a wavelength effective to activate the photosensitizing agent, such that the membrane of said intracellular compartment is disrupted, releasing said molecule into the cytosol of the cell, without killing the cell,

wherein, said released antigenic molecule, or a part thereof, is subsequently presented on the surface of said cell.

3. The method of claim 1 or claim 2 wherein the antigenic molecule is a molecule capable of stimulating an immune response.

4. The method of claim 3 wherein the antigenic molecule is a vaccine antigen or vaccine component.

5. The method of any of the preceding claims wherein the antigenic molecule is a peptide.

6. The method of any of the preceding claims wherein the cell is an antigen presenting cell selected from the

group comprising lymphocytes, dendritic cells, macrophages and cancer cells.

7. The method of any of the preceding claims wherein the photosensitizing agent is selected from the group comprising porphyrins, phthalocyanines, purpurins, chlorins, benzoporphyrins naphthalocyanines, cationic dyes, tetracyclines and lysomotropic weak bases or derivatives thereof.

8. The method of claim 7 wherein the photosensitizing agent is TPPS₄, TPPS_{2a} or AlPcS_{2a}.

9. The method of any of the preceding claims wherein the antigenic molecule and/or photosensitizing agent is bound to one or more targeting agents or carrier molecules.

10. The method of any of the preceding claims wherein said method is carried out *in vitro* or *in vivo*.

11. The method of any of the preceding claims wherein the antigenic presentation results in the stimulation of an immune response.

12. A method of vaccination comprising the method as defined in any one of the preceding claims.

13. A composition for use in expressing an antigenic molecule or a part thereof on the surface of a cell, said composition comprising an antigenic molecule as defined in any one of claims 1 to 5 or claim 9 and a photosensitizing agent as defined in any one of claims 2 or 7 to 9.

14. A cell expressing an antigenic molecule, or a part thereof, on its surface, or a population thereof, which

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cell is obtainable by a method as defined in any one of claims 1 to 12.

15. A composition as defined in claim 13 or a cell
5 population as defined in claim 14 for use in therapy.

16. Use of an antigenic molecule and a photosensitizing
agent as defined in the preceding claims in the
preparation of a medicament for use in expressing said
10 antigenic molecule or a part thereof on the surface of a
cell to stimulate an immune response or for stimulating
CTLs.

17. Use of a cell population as defined in claim 14 for
15 the preparation of a medicament for stimulating an
immune response or for stimulating CTLs.

18. Use as claimed in claim 16 or claim 17 wherein said
medicament is used in vaccination.

19. Use as claimed in any one of claims 16 to 18
20 wherein said medicament is used in the treatment of
viral diseases, cancer or multiple sclerosis.

20. A product comprising an antigenic molecule as
25 defined in any one of claims 1 to 5 or claim 9 and a
photosensitizing agent as defined in any one of claims 2
or 7 to 9 as a combined preparation for simultaneous,
separate or sequential use in expressing said antigenic
30 molecule or a part thereof on the surface of a cell.

21. A kit for use in expressing an antigenic molecule
or a part thereof on the surface of a cell, said kit
comprising
35 a first container containing said antigenic
molecule as defined in any one of claims 1 to 5 or claim
9; and

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a second container containing a photosensitizing agent as defined in any one of claims 2 or 7 to 9.

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ABSTRACT

The present invention provides a method of
5 expressing an antigenic molecule or a part thereof on
the surface of an antigen-presenting cell, said method
comprising introducing a molecule into the cell cytosol
by photochemical internalisation, wherein said molecule,
or a part thereof, is subsequently presented on the
10 surface of said cell. Methods of vaccination comprising
this method, together with compositions comprising said
cells and uses involving said cells expressing antigenic
molecules are also provided.

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Washington, D.C. 20231

Printed Name Shawn Hipe

Signature [Signature]

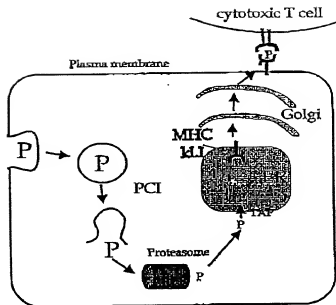


FIGURE 1

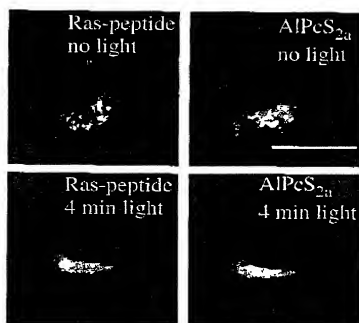


FIGURE 2

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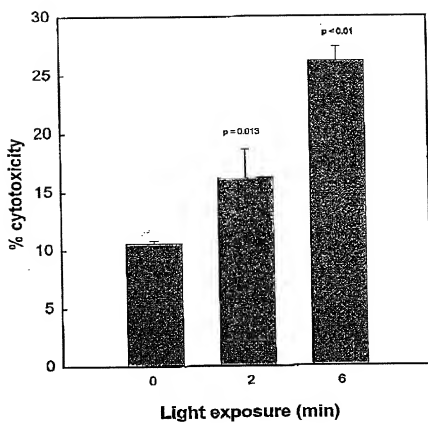


Figure 3

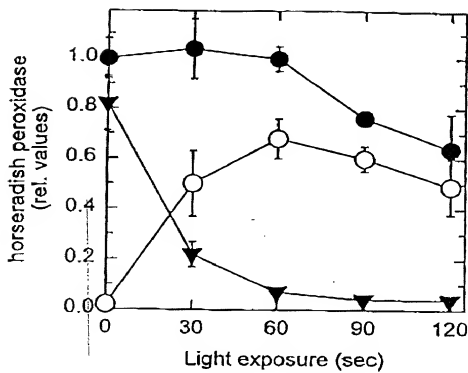


FIGURE 4

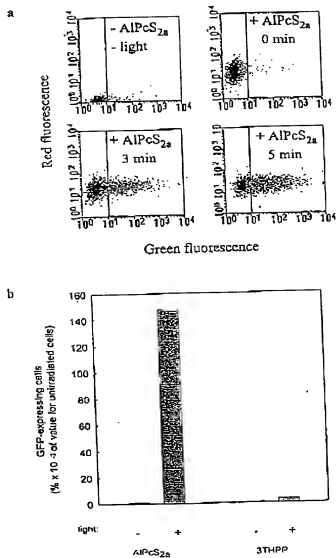


FIGURE 5

SCHWEGMAN, LUNDBERG, WOESSNER & KLUTH, P.A.

United States Patent Application

COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor I hereby declare that: my residence, post office address and citizenship are as stated below next to my name; that

I verily believe I am the original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled: **METHOD OF VACCINATION**.

The specification of which is attached hereto.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with 37 C.F.R. § 1.56 (attached hereto). I also acknowledge my duty to disclose all information known to be material to patentability which became available between a filing date of a prior application and the national or PCT international filing date in the event this is a Continuation-In-Part application in accordance with 37 C.F.R. § 1.63(e).

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on the basis of which priority is claimed:

Application Number	Country	Day/Month/Year Filed
9905911.5	United Kingdom	15/03/1999

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below:

No such claim for priority is being made at this time.

I hereby claim the benefit under 35 U.S.C. § 120 or 365© of any United States and PCT international application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT international application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose material information as defined in 37 C.F.R. § 1.56(a) which became available between the filing date of the prior application and the national or PCT international filing date of this application:

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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Date: _____

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Citizenship:
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(a) A patent by its very nature is affected with a public interest. The public interest is best served, and the most effective patent examination occurs when, at the time an application is being examined, the Office is aware of and evaluates the teachings of all information material to patentability. Each individual associated with the filing and prosecution of a patent application has a duty of candor and good faith in dealing with the Office, which includes a duty to disclose to the Office all information known to that individual to be material to patentability as defined in this section. The duty to disclose information exists with respect to each pending claim until the claim is canceled or withdrawn from consideration, or the application becomes abandoned. Information material to the patentability of a claim that is canceled or withdrawn from consideration need not be submitted if the information is not material to the patentability of any claim remaining under consideration in the application. There is no duty to submit information which is not material to the patentability of any existing claim. The duty to disclose all information known to be material to patentability is deemed to be satisfied if all information known to be material to patentability of any claim issued in a patent was cited by the Office or submitted to the Office in the manner prescribed by §§ 1.97(b)-(d) and 1.98. However, no patent will be granted on an application in connection with which fraud on the Office was practiced or attempted or the duty of disclosure was violated through bad faith or intentional misconduct. The Office encourages applicants to carefully examine:

- (1) prior art cited in search reports of a foreign patent office in a counterpart application, and
 - (2) the closest information over which individuals associated with the filing or prosecution of a patent application believe any pending claim patentably defines, to make sure that any material information contained therein is disclosed to the Office.
- (b) Under this section, information is material to patentability when it is not cumulative to information already of record or being made of record in the application, and
- (1) It establishes, by itself or in combination with other information, a prima facie case of unpatentability of a claim; or
 - (2) It refutes, or is inconsistent with, a position the applicant takes in:
 - (i) Opposing an argument of unpatentability relied on by the Office, or
 - (ii) Asserting an argument of patentability.

A prima facie case of unpatentability is established when the information compels a conclusion that a claim is unpatentable under the preponderance of evidence, burden-of-proof standard, giving each term in the claim its broadest reasonable construction consistent with the specification, and before any consideration is given to evidence which may be submitted in an attempt to establish a contrary conclusion of patentability.

- (c) Individuals associated with the filing or prosecution of a patent application within the meaning of this section are:
- (1) Each inventor named in the application;
 - (2) Each attorney or agent who prepares or prosecutes the application; and
 - (3) Every other person who is substantively involved in the preparation or prosecution of the application and who is associated with the inventor, with the assignee or with anyone to whom there is an obligation to assign the application.
- (d) Individuals other than the attorney, agent or inventor may comply with this section by disclosing information to the attorney, agent, or inventor.